		ent 1.				Date 18/2191
. (]	FACE	SHEET) ETHICAL	REVI	EW CO	MMETTE	E, ICDDR, B.
D.	rincia	•				
	Tuci	al investigator by, San	<u> </u>	-sako	, Traiı	nee Investigator (if any)
A	pplice	ition No. PCC/005/c	10/(Re	vised	) Suppo	orting Agency (if Non-ICDDR,B)
Ti	itle o	of Study Study on the	Preiv	A. 60 (	J Proje	ect status:
P	برطياه	lateral are lable to a top	41.4	·	( )	New Study
		lobactus Jeffer and c	Leve	Und	( )	Continuation with change
_1	ei'h	motile and hon-walle	Vasi	ants.	( )	No change (do not fill out rest of form)
Ci	rcle	the appropriate answer	to eac	h of	the fe	ollowing (If Not Applicable write NA).
1.	Sou	Fermorout.	eo cac	,11 O1	5.	Will signed concent form by
	(a)		(Yes	) No	•	Will signed consent form be required: (a) From subjects  Yes No
	(p)		Yes			(a) From subjects Yes No (b) From parent or guardian
	(c)					(if subjects are minors) Yes No
-		under guardianship	Yes	No	6.	Will precautions be taken to protect
2.	νoe	s the study involve:				anonymity of subjects Yes No
	(a)			~~	7.	Check documents being submitted herewith
	(b)	subjects	Yes	F-2-6	-	Committee:
	(c)		Yes	(No)		Umbrella proposal - Initially submit
	(c)	Psychological risks to subjects				overview (all other requirements wil
	(d)	Discomfort to subjects	Yes	) — ·		be submitted with individual studies
	(e)	Invasion of privacy		\\		Protocol (Required)
	(£)	Disclosure of informa-	Yes	(No)	•	Abstract Summary (Required)
		tion damaging to sub-				Statement given or read to subjects
		ject or others	Yes	(No)		nature of study, risks, types of que
3.	Does	the study involve:	103	(III)		TORS TO be asked, and right to refus
	(a)	Use of records, (hosp-	•			to participate or withdraw (Required)
		ital, medical, death.				informed consent form for subjects
		birth or other)	Yes	No		Informed consent form for parent or guardian
	(b)	Use of fetal tissue or		- 1		
	(-)	abortus	Yes	(No)		Procedure for maintaining confidential
	(c)	Use of organs or body	~····	$\overline{}$		
4.	A	fluids	(Yes)	) No		Questionnaire or interview schedule *  * If the final instrument is not completed
٠,	(a)	subjects clearly informed	ed abo	ut:		prior to review, the following informati
	(a)	Nature and purposes of				should be included in the abstract summa
	(b)	Study	(Yes)	No		1. A description of the areas to be
	(0)	Procedures to be	$\smile$			covered in the questionnaire or
		followed including alternatives used				interview which could be considered
	(c)	Physical risks		No		either sensitive or which would
	(d)	Sensitive questions	Yes	No		constitute an invasion of privacy
	(e)	Benefits to be derived	Yes	No		2. Examples of the type of specific
	. 9.	OC GOTTAGO	Yes	No		questions to be saled

privacy. or the type of specific questions to be asked in the sensiti areas.

An indication as to when the question naire will be presented to the Cttee for review.

We agree to obtain approval of the Ethical Review Committee for any changes involving the right/s and welfare of subjects before making such change.

Yes

Yes

No

No

Principal Investigator

Right to refuse to

draw from study

of data

participate or to with-

Compensation 6/or treatment where there are risks or privacy is involved in

any particular procedure Yes No

Confidential handling

(<del>‡</del>)

(g)

(h)

Trainee

## APPLICATION FOR PROJECT GRANT

- 1. TITLE OF THE PROJECT
- Study on the virulence of Campylobacter jejuni and C. coli and their motile and non-motile variants
- 2. PRINCIPLE INVESTIGATOR
- Dr. Samir K. Saha ()() u Dhaka Shishu Hospital (DSH)

3. COINVESTIGATORS

a) Dr. Waqar A. Khan Dhaka Shishu Hospital

Men

b) Dr. M. John Albert ICDDR,B

4. CONSULTANT

Prof. M. S. Akbar Dhaka Shishu Hospital

5. STARTING DATE

October 01, 1990

- 6. COMPLETION DATE
- : September 30, 1991
- 7. TOTAL BUDGET REQUIRED
- US\$ 5,185

8. APPROVALS

Director

Dhaka Shishu Hospital

3. 10.000

Head

Laboratory Sciences Division

ICDDR.B

## 9. SUMMARY

Recently, a small, sensitive and reproducible animal has been developed by the principal investigator to test the enterotoxicity of Campylobacter jejuni (Saha et al., 1988) which, subsequently, was also found to be suitable to study the colonisation of the organism and the histopathological changes caused by the organism (Saha and Sanyal, 1990; S.K. Saha, Ph.D. thesis, 1990). Use of this animal model for further detailed study of Campylobacter should resolve the existing discrepancies regarding some of the pathogenesis of C. jejuni and C. coli and the role of flagella in the enteropathogenicity of C. jejuni/coli.

This proposal aims to study the prevalence and comparative virulence of *C. jejuni* and *C. coli* and their virulence patients and in experimental animal model. The study also aims to the determination of role of flagella in virulence in rat ileal loop model by using the isogenic non-motile Campylobacter jejuni/coli. variants of Virulence in patients will be studied on the basis of the macroscopic and microscopic appearance of faeces, duration of diarrhoea, abdominal pain, vomiting, dehydration, temperature. haemotological values, haemolytic process, etc. Virulence in animal model will be studied on the basis of quantitative assay of enterotoxin, colonising ability, infective dose and relative histopathological changes.

The work will help to identify the prevalent species of Campylobacter in Bangladeshi children and to understand the relative virulence of *C. jejuni* and *C. coli* and their motile and non-motile forms, in patients and experimental animal model.

Study on motile and non-motile variants will be of paramount significance to understand the role of flagella in virulence and whether the flagellin epitope(s) should be considered as potential targets against which the host antibodies has to be directed for immunoprophylactic measures against Campylobacter enteritis.

## 10. AIMS OF THE PROJECT

- a) General aims
  - Determination of prevalence of Campylobacter jejuni and C. coli in diarrhoeal and nondiarrhoeal patients.
  - 2) Study on the relative level virulence of C. jejuni/C. coli in clinical settings and in experimental animal model. And the comparative virulence of motile and non-motile strains in animal model.

- b) Specific aims
  - Campylobacter strains will be isolated and identified to see the prevalence of C. jejuni and C. coli and in diarrhoeal and non-diarrhoeal patients.
  - 2) Comparative virulence of the strains in patients will be determined on the basis of the following clinical symptoms:
    - i) macroscopic and microscopic appearance of stool
    - ii) duration of diarrhoea
    - iii) abdominal pain
      - iv) vomiting
        - v) dehydration
      - vi) temperature
  - 3) Comparative virulence of the strains in rat ileal loop model will be determined on the basis of the following criteria:
    - i) comparative infective dose (cfu)
    - ii) ability to cause the histopathological changes in rat gut
    - iii) ability to colonise the rat gut
  - The non-motile isolates/variants of *C. jejuni* and *C. coli* will be serially passaged through rat gut to see whether the strains will revert to motile form.

## 11. SIGNIFICANCE

The above mentioned works on Campylobacter will help to identify the predominant Campylobacter species in Bangladeshi children and the comparative virulence of C. jejuni and C. coli in causing enteritis.

The detailed comparative study on motile and non-motile variants will facilitate better understanding of pathogenesis of Campylobacter.

Finally, the study shall have a paramount significance to determine the role of flagella in virulence and whether the flagellin epitopes should be considered as the potential targets against which the host antibodies have to be directed for immunoprophylactic measures against *Campylobacter* enteritis.

## 11. ETHICAL IMPLICATION

No major ethical issues are involved.

### 12. BACKGROUND

Campylobacter jejuni/coli is now well-established as one of the commonest causes of enteritis in man (Walker et al., 1986). The two species, C. jejuni and C. coli, are very closely related. Studies have shown that the prevalence of the species in man is not the same at different parts of the world (Skirrow, 1984; Georgecurbot et al., 1986).

Furthermore, two studies, from developed countries, regarding the virulence of these two species in human being is in contrast to each other, one group claimed *C. jejuni* as more virulent and another group, *C. coli* and *jejuni* having the same degree of virulenceas (Papovic-Uroic *et al.*, 1988; Figura and Guglielment, 1988). These studies were only clinical-symptom-oriented. However, there is no such data from the developing countries.

The exact mechanism of the pathogenicity of Campylobacter is both clinical obscurity. There are in experimental evidences to suggest that invasion (Butzler and Skirrow, 1979) and/or production of enterotoxin (Ruiz-Palacios et al., 1983; Saha et al., 1988; Saha and Sanyal, 1990) play an important role in the pathogenesis of Campylobacter. In certain established enteropathogens, such Vibrio cholerae (Attridge and Rowley, 1983) Salmonella typhimurium (Carsiotis et al., 1984), motility has been established as an important virulence factor. reports on the possible role of motility in the virulence of Campylobacter are scanty. Two groups of workers (Morooka et al., 1985; Newell et al., 1985) observed a significant role of motility for the pathogenesis of the organism but another group (Field et al., 1986) failed to correlate virulence of Campylobacter with its motility. Two available reports regarding the reversion of non-motile C. jejuni/coli strains to a motile phenotype in vivo is also in contrast to each other (Caldwell et al., 1985; Field et al., 1986).

All these discrepancies in the studies on pathogenicity of Campylobacter could be attributed to the absence of a suitable animal model (Walker et al., 1986). However, recently we have reported that an iteal loop model using C-F rats (Saha et al., 1988, Saha and Sanyal, 1990), is excellent in sensitivity and reproducibility and can be used further for studying the pathogenicity of C. jejuni and C. coli.

### 13. RESEARCH PLAN

Rectal swab/stool samples will be collected from diarrhoeal (watery/bloody) patients of age groups of 3 months to five years and age-sex matched non-diarrhoeal controls admitted in the Dhaka Shishu Hospital (DSH). Campylobacter strains be isolated on Campy-BAP (Blaser et al., will following the method of Saha and Sanyal (1989) and will identified by the method of Skirrow and Benjamin (1980). simultaneously cultured specimens will be MacConkey, Salmonella-Shigella and TCBS agar to exclude presence of other enteropathogenic bacteria. The iodine preparation of stool specimens will be microscopically examined to exclude the presence of parasite(s). Presence of rotavirus, ETEC (Svennerholm et al., 1986) and EPEC (Albert et al., submitted) will be excluded by the ELISA test.

#### Clinical

Fifty cases, from which Campylohacter jejuni/coli will be isolated as a sole pathogen, will be studied on the basis of the clinical features, such as watery diarrhoea, bloody mucoid diarrhoea, mucoid diarrhoea without blood, abdominal pain, vomiting, dehydration, peak temperature, white blood cell count, etc.

Blood specimens routinely referred to our pathology laboratory will be used to do the haematological tests mentioned in the proposal.

### Bacteriological

The non-motile variants of 12 representative motile isolates of Campylobacter (6 C. jejuni and 6 C. coli) will be screened by using 0.5% agar plates (Field et al. 1986) just after isolation. The non-motile strains of C. jejuni and C. coli will be further confirmed by flagellar staining (Leifson's method).

## **EXPERIMENTAL**

## Enterotoxins

Six Campylobacter jejuni and six C. coli strains (both their motile and non-motile isogenic variants), with least possile subculture, will be tested in rat ileal loop model (Saha et al., 1988). The comparative enterotoxicity of the strains will be interpreted by measuring the ml of fluid accumulated per cm of rat gut.

#### Infective dose

To determine any differences in the infective doses [the minimum number of bacteria that cause full-blown reaction (0.5%)] of C, jejuni and C, coli and their motile and non-motile variants, graded doses (cfu) of the strains will be injected into rat ileal loops.

#### Reversion

Non-motile strains will be serially passaged through the rat gut by the method described earlier (Saha et al., 1988), and any change in their motility will be tested after each passage. Observation of any reversion of a non-motile strain to a motile one, on soft agar plates, will be confirmed by flagellar staining and.

#### Colonisation/Adherence

Comparative ability of motile and non-motile strains of Campylobacter to colonise/adhere the rat gut will be evaluated by determining the cfu of the organism in mucus scrappings of rat gut after inoculation with same number of bacteria of both, motile and non-motile, variants. Lumen of the gut will be gently washed with normal saline before taking the mucosal scrappings.

## Histopathology

Neighbouring rat ileal loops, inoculated with similar cfu of motile and non-motile variants, will be processed (after

18 hr of inoculation) for histopathological study. In brief, the microtome sections of respective gut loop will be stained by Haemotoxylin and Eosin method (Luna, 1968). The stained sections of the loops with motile/non-motile variants of *C. jejuni/coli* will be comparatively examined for any change in crypt:villi ratio, cellular infiltration of epithelium, sloughing of mucosal cells, etc.

#### 14. BIBIOLOGRAPHY

- 1) Attridege SR and Rowley D. (1983) The role of the flagellum in the adherence of *Vibrio cholerae*. J. Infect. Dis. 147:864-872.
- 2) Blaser MJ, Berkowitz ID, LaForce FM, Dravens J, Reller LB and Wang WL. (1979) Campylobacter enteritis: Clinical and epidemiologic features. Ann. Intern. Med. 91:179-185.
- 3) Butzler JP and Shirrow MB. (1979) Campylobacteri enteritis. Clin. Gastrol. 8:737-765.
- 4) Caldwell MB, Walker RI, Stewart SD and Rogers JE. (1983)
  Simple adult rabbit model for Campylobacter jejuni
  enteritis. Infect. Immun. 42:1176-1182.
- 5) Carsiotis M, Weinstein DL, Karch H, Holder IA and O'Brien AD. (1984) Flagella of Salmonella typhimurium are a virulence factor in infected C57BL/6J mice. Infect. Immun. 46:814-818.

- 12) Ruiz-Palacios GM, Torres J, Torres NI, Escamilla E, Ruiz-Palacios B and Tamaya J. (1983) Cholera-like enterotoxin produced by Campylobacter jejuni. Characterisation and clinical significance. Lancet ii:250-252.
- of chicken isolates of Campylobacter jejuni in ligated ileal loops of rats. J. Med. Microbiol. 26(2):87-91.
- 14) Saha SK and Sanyal SC. (1989) Better growth of Campylobacter jejuni using simple Fortner's principle and candle extinction jar. Indian'J. Med. Res. 89:24-27.
- 15) Saha SK and Sanyal SC. (1990) Production and characterisation of Campylobacter jejuni enterotoxin in a synthetic medium and its assay in rat ileal loops. FEMS Microbiol. Lett. 67:333-338.
- 16) Sanyal SC, Islam KMN, Neogi PKB, Islam M, Speelman P and Huq MI. (1984) Campylobacter jejuni diarrhoea model in infant children. Infect. Immun. 43:931-936.
- 17) Skirrow MB and Benjamin J. (1980) A human strain of Campylobacter fetus sub sp. intestinalis grown at  $42^{\circ}$ C. J. Clin. Pathol. 33:603-604.
- 18) Skirrow MB. (1984) Campylobacter infections of man.

  Easmon CSF and Jeljaszewicz J (eds.). In: Medical

  Microbiology, vol. 4, pp.105-141.

- 6) Field LH, Headley VL, Underwood JL, Payne SM and Berry LJ.

  (1986) The chicken embryo as a model of Campylobacter invasion: Comparative virulence of human isolates of Campylobacter jejuni and Campylobacter coli. Infect. Immun.

  54:118-125.
- 7) Figura N and Guglielmetti P. (1988) Clinical characteristics of Campylobacter jejuni and C. coli enteritis. Lancet i:942-943.
- (8) Georges-Courbot MC, Baya C, Berand AM, Mennier DMY and Georges AJ. (1986) Distribution and serotypes of Campylobacter jejuni and Campylobacter coli in enteric Campylobacter strains isolated from children in the Central African Republic. J. Clin. Microbiol. 23:592-594.
- Morooka T, Umeda A and Amako K. (1985) Motility as an intestinal colonization factor for Campylobacter jejuni. J.
   Gen. Microbiol. 131:1973-1980.
- 10) Newell DG, McBride H, Saunders F, Dehele Y and Pearson AD.

  (1985) The virulence of clinical and environmental isolates of Campylobacter jejuni. J. Hyg. 94:45-54.
- 11) Papovic-Uroic T, Ginajuicki B, Kalemic S and Vodopija J.

  (1988) Clinical comparison of Campylobacter jejuni and

  C. coli diarrhoea. Lancet i:176-177.

- 19) Svennerholm AM, Wikston M, Lindblad M and Holgren J. (1986)

  Monoclonal antibodies against *E. coli* heat-stable enterotoxin (ST) and their use in diagnostic ganglioside GM<sub>1</sub> enzyme-linked immunosorbent assay. J. Clin. Microbiol. 24:585-590.
- 20) Walker RI, Cadwell MB, Lee EC, Guerny P, Trust TJ and Ruiz-Palacios GM. (1986). Pathophysiology of *Campylobacter* enteritis. Microbiol. Rev. 50:81-94.

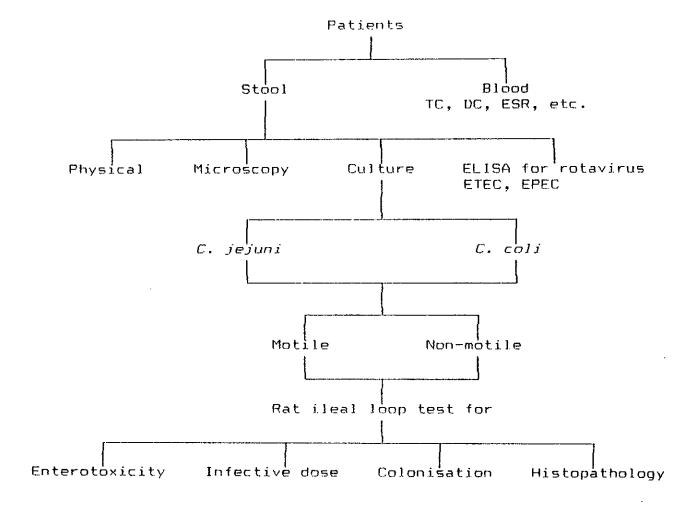
## 15. PUBLICATIONS OF PRINCIPAL INVESTIGATOR

- Saha SK, Aziz KMS and Khan WA. (1985) Escherichia coli: a
  possible source of skin infection in Bangladesh (Abstract).
   4th Annual Conference of BSM, JCDDR, B, Dhaka.
- 2) Akbar MS, Naila Khan and Saha SK. (1986) Evaluation of chloramphenical and co-trimoxazale in the treatment of enteric fever. Bangladesh J Child Healt 10(1):11-14.
- 3) Naila Khan, Saha SK and Akbar MS. (1986) Comparative efficacy of blood, urine, stool and bone marrow cultures for isolation of Salmonella typhi in typhoid fever. J. Bangladesh Col. Phys. Surg. 3(2):10-13.
- and Ahsan CR. (1987)4) SC. Saha S, Saha SK Sanyal Immunologic and genetic relationship between enterotoxins. Р. toxin research. and In: Progress in AGDOM Gopalakrishnakone and CK Tan (eds), pp.569-678.

- Sanyal SC, Sahak SK, Shukla BN, Saha S, Singh NP and Agarwal RK. (1987). Human, poultry and water isolates of Campylobacter jejuni produce CT-like toxin in Charles-Foster strain of rats. In: Proceedings of First Asian Congress on Anaerobic Bacteria in Health and Disease. A. Mehta and A.N. Kochar (eds), pp.44-52.
- 6) Saha SK, Singh NP and Sanyal SC. (1988) Enterotoxicity of chicken isolates of *Campylobacter jejuni* in ligated ileal loops of rats. J. Med. Microbiol. 26(2):87-91.
- 7) Sanyal SC, Saha SK and Shukla BN. (1988) Clinical, environmental and chicken isolates of Campylobacter jejuni produce CT-like enterotoxin (Abstract WeP 4-16), XIIth International Congress for Tropical Medicine and Malaria, Amsterdam, The Netherlands. P.A. Kager and A.M. Polderman (eds.), pp.215.
- 8) Saha SK. (1989) Use of beta-lactamase inhibitors: A noble approach to bring back the older beta-lactams. Dhaka Shishu (Children) Hospital Journal, 5:35-37.
  - 9) Waqar AK, Saha SK, Akbar MS and Hoq MS. (1989). Aetiology of meningitis in Bangladeshi children. Dhaka Shishu (Children) Hospital Journal, 5:5-9.
- 10) Saha SK and Sanyal SC. (1989) Better growth of Campylobacter jejuni using simple Fortner's principle and candle extinction jar. Indian J. Med. Res. 89:24-27.

- 11) Saha SK and Sanyal SC. (1990) Production and characterisation of *Campylobacter* enterotoxin in a synthetic medium and its assay in rat ileal loops. FEMS Microbiol. Lett. 67:333-338.
- 12) Saha SK and Sanyal SC. (1990) Improved medium for the preservation of a *Campylobacter jejuni*. Indian J. Med. Res. (in press).

## 16. FLOW CHART



## 17. ITEMIZED SPECIFIC TASKS FOR EACH LISTED INVESTIGATOR

- a) Dr. Samir K. Saha Will supervise the whole work and will do the animal experiment
- b) Dr. Waqar A. Khan Will examine the histopathological and haematological slides
- Dr. M. John Albert Wj ] ] supervise and render expert c) the time o f animal help аt experiments and ELISA tests at ICDDR.B
- d) The data for clinical evaluation will be collected from the hospital record files of the respective patients and Prof. M.S. Akbar will be available for any extra information and/or interpretation.

## 18. COLLABORATION

- 1) All the work will be done at Dhaka Shishu Hospital (DSH), except the ileal loop tests and ELISA, which will be done at ICDDR.B.
- 2) Campylobacter isolates will be preserved at  $-70^{\circ}$ C at ICDDR,B.
- 3) If needed, some Campylobacter strains will be obtained from ICDDR.B through proper channel.

## 19. FACILITIES AVAILABLE AT DHAKA SHISHU HOSPITAL

Dhaka Shishu Hospital, the only children's hospital at national level of Bangladesh, has 350 beds. Isolation of *Campylobacter* and other enteropathogens can be done at DSH. The PI has got the supply of antibiotic supplements and chemicals for the isolation and identification of *Campylobacter*, which he brought with him from India.

tests will be done at 1CDDR.B. but the Rat ileal loop colonisation and histopathological studies will be done at Any reversion of non-motile Campylobacter to a motile phenotype, after animal passage, will be tested in DSH. All the facilities, , machine (Reichert-Jung), including microtome to dσ the histopathological studies are available at DSH.

# 20. BUDGET

W.

a)	Personnel

Name,	Time input 	Cost/annum	
Dr. Samir K. Saha, Ph.D. Principal Investigator	30%	Tk. 36,000	
Dr. Waqar A. Khan, M.Phil. Coinvestigator	10%	12,000	
Dr. M. John Albert, Ph.D. Coinvestigator	5%	-	
Research Officer (MBBS/M.Phil/M.Sc.) (to be recruited)	100%	48,000	
Laboratory Assistant	100%	24,000	120,000
Consultant:		·	
Prof. M. S. Akbar, MRCP Will be consulted when re	- equired	-	-
Supplies			•
Média and glasswares	•	Tk. 18,000	
Chemicals and reagents		7,000	
Animals (rats)	,	7,000	
ELISA tests		7,000	39,000
Equipment			
Refrigerator - one			20,000
Overhead/contingencies			5,000
PROJECT COST		Tk.	184,000